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# Pharmacological insights into modified citrus pectin: A promising therapeutic potential with anticancer effect

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**Running Title:** Anticancer and pleiotropic effects of modified citrus pectin

#### **Abstract**

Modified citrus pectin (MCP) is a naturally soluble dietary fiber derived from citrus pectin that inhibits galectin-3 (Gal-3), a proinflammatory, profibrotic, and prometastatic regulatory protein. Interestingly, the anticancer activity of MCP against multiple cancers, such as colon, prostate, urinary bladder, hemangiosarcoma, and breast cancers, has been demonstrated. It could be attributed to its inhibitory effect on cancer cell growth, prevention of metastasis, and induction of cancer cell apoptosis. In addition, MCP presented protective effects against organ damage in different disease models, including cardioprotective, neuroprotective, renoprotective, hepatoprotective effects. This could be ascribed to its antioxidant, anti-inflammatory, anti-apoptotic, and antifibrotic effects. Further, immunomodulatory, detoxifying, antimicrobial, and chondroprotective effects have also been demonstrated with MCP. It is available in the market as a regarded safe dietary supplement due to its healthpromoting effects. This review involves the interplay between cancer, Gal-3, and MCP, as well as the beneficial impacts of MCP in several models of organ damage.

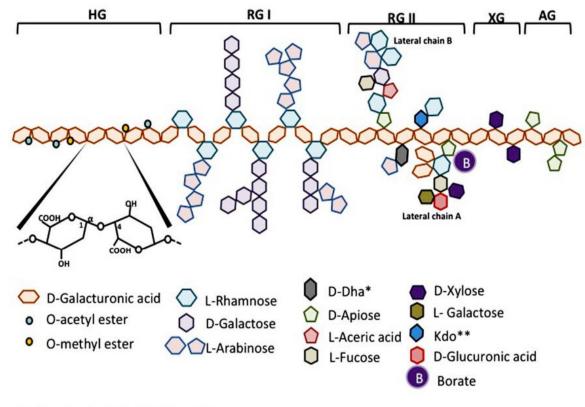
Keywords: modified citrus pectin, galectin-3, cancer, organ damage

#### 1. Introduction

Citrus pectin was modified to enhance biological activities and introduced as modified citrus pectin (MCP). Citrus fiber is obtained from citrus fruits' albedo and membrane parts.<sup>1</sup> Pectin is а complex heteropolysaccharide that consists mainly of two parts: a linear part named the smooth region (homogalacturonan (HG)) and a branched part named the hairy region (rhamnogalacturonan I (RG-I) and rhamnogalacturonan II (RG-II)). This is demonstrated in Figure 1. Pectin occurs primarily as a protopectin characterized by high molecular weight (Mw), gel-like low solubility, structure, and constraining its intestinal absorption and utility in certain fields.<sup>2</sup> Initially, modifying citrus pectin was done at certain temperatures and different pH levels, leading to low Mw and less esterified product, facilitating intestinal absorption into the blood circulation.<sup>3,4</sup> MCP is marketed as PectaSol, prepared by pH-controlled enzymatic treatment,

and is available in capsule or powder form. The recommended dose by the manufacturer is fifteen grams to be divided 2-3 times per day and taken with water or juice. 5,6

The Food and Drug Administration (FDA) of the United States classifies MCP as a generally regarded safe dietary supplement.<sup>7</sup> It is sold as a dietary supplement due to its healthpromoting effects, such as heavy metal elimination, antioxidant activity, antiinflammatory, and hypocholesterolemic effects.8-10 Moreover, MCP exhibited immunomodulatory effects increasing proinflammatory cytokines.<sup>11</sup> Further, the anticancer effects of MCP were demonstrated in different preclinical and clinical studies. It inhibits cancer cell growth and prevents metastasis while inducing cancer cell apoptosis.5,12 Most of the research on MCP focuses on its galectin-3 antagonism.



<sup>\*</sup>D-Dha = 3-deoxy-D-lyxo-2-heptulosaric acid

**Fig. 1** Schematic illustration of pectin's structure. AG: Arabinogalactan, HG: Homogalacturonan, RG: Rhamnogalacturonan, XG: Xylogalacturonan. Taken, with permission under Creative Commons Attribution License (CC BY), from Anti-cancer activities of pH- or heat-modified pectin.<sup>13</sup>

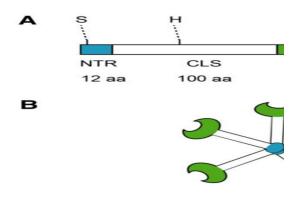
#### 2. Galectin-3

Galectin-3 (Gal-3) is a multifunctional  $\beta$ -galactoside binding lectin. It is primarily synthesized and secreted by macrophages, eosinophils, and mast cells. It binds to ligands having a  $\beta$ -galactoside structure through its carbohydrate-recognition domain

(CRD). Diverse glycosylated matrix proteins, including integrins, fibronectin, and laminin, are among the numerous Gal-3 ligands that have been recognized. 14,15 It is a chimera-type galectin indicated by a monomeric structure that comprises a short N-

<sup>\*\*</sup>Kdo = 3-deoxy-D-manno-2-octulosonic acid

terminal domain, a C-terminal domain, and an intermediate repeat domain of proline-glycine-alanine-tyrosine<sup>16-18</sup> as shown in Figure 2. Gal-3 has a complicated mechanism of action and remains empirical until now. A possible explanation is that it interacts with different proteins many in the extracellular matrix, inside the cell, at the cell membrane, and in biological Consequently, it has been fluids. documented as participating in physiological and numerous pathological events. 19-22 It is engaged in



**Fig. 2** Structure of galectin-3. (A) Galectin-3 protein structure consists of the carbohydrate recognition domain (CRD) of 130 amino acids (aa), which comprises the C-terminal and contains

The most significant clinical challenge related to cancer is metastasis.

various biological events, including cell migration. adhesion. apoptosis, angiogenesis, and inflammation. Its pivotal role in tissue fibrosis and inflammation has been documented. 23,24 While Gal-3 has traditionally been viewed as a biomarker linked to disease,<sup>25</sup> recent research has clearly shown its significant role therapeutic target for various fibrotic and inflammatory conditions.<sup>26</sup> It is worth noting that the contribution of Gal-3 to the progression and metastasis of cancer has been documented.<sup>27-30</sup>

the anti-death motif or Asp-Trp-Gly-Arg (NWGR). The N-terminal Domain (NTD), which has an N-terminal region of 12 amino acids and contains a serine phosphorylation site. (S) Pentameric structure of Gal-3. Taken permission under Creative Commons Attribution License (CC BY) from Galectin-3 in Atrial Fibrillation: Mechanisms Therapeutic and Implications.<sup>31</sup>

# 3. Interplay between cancer, Gal-3, and MCP

Metastasis is the dissemination of cancer cells from the primary tumor

growth to distant tissues and organs. It is the leading cause of morbidity and death linked to cancer. Regarding the metastatic cascade. Gal-3 and subsequently MCP modulate different rate-limiting steps. MCP is now recognized as an up-and-coming antimetastatic agent.<sup>12</sup> The initial step undertaken by the cancer cells, after escaping from the primary tumor and undergoing intravasation. to overcome apoptosis linked to loss of anchorage (anoikis). Gal-3 safeguards cancer cells from anoikis. 32,33 It induces cell cycle arrest at the late G1 phase, which is an anoikis-resistant point.<sup>32</sup> MCP induced apoptosis of human JCA-1 cells prostatic through downregulating cyclin B and cdc2,<sup>34</sup> which may accumulate neoplastic cells in the G2/M phase, consequently inducing apoptosis.<sup>12</sup>

The following rate-limiting step of metastasis entails cancer cell arrest in the microvasculature of the distant organ. Gal-3 has been demonstrated to facilitate the adhesion of metastatic cells to the endothelium. 35-38

Conversely, MCP inhibited their adhesion to endothelium and their homotypic aggregation, which is associated with the arrest of metastatic cells in distant organs and metastatic deposit formation intravascularly.<sup>4,39-41</sup>

Upon infiltrating the microvessels of the target organ, the cancer cells may proliferation either undergo intravascularly till the metastatic tumor surpasses the blood vessel and invades the parenchyma of the distant organ<sup>42</sup> or extravasate before commencing secondary tumor growth. Invasive propensity encompasses a series of interactions between extracellular matrix proteins, related to target organ stroma and basement membrane, and tumor cells mediated by Gal-3.43 MCP inhibited these Gal-3-mediated interactions. Citrus pectin polysaccharides reduced the invasion of human endothelial cells through the Matrigel dose-dependently, 40 as well as that of human metastatic buccal carcinoma and MDA-MB-231 human breast carcinoma metastatic cells.<sup>44</sup>

Following the first parking in distant organs and extravasation, almost all cancer cells experience apoptosis triggered by different factors, and less 2% survive than and micrometastasis.45 Consequently, one of the most critical rate-limiting steps influencing metastasis efficacy is the clonogenic survival of early metastatic colonies. It was described that Gal-3 is important in neoplastic cell clonogenic survival through its anti-apoptotic effects, working on the mitochondrial apoptosis pathways. 46-48 Some reports indicated that MCP-induced Gal-3 blockade could antagonize the antiapoptotic effects of Gal-3, hence decreasing cancer cells' clonogenic survival.<sup>48</sup> Accordingly, MCP hindered the hemangiosarcoma cells' clonogenic survival in a dose-dependent way, increasing the apoptosis of tumor cells.49

As micrometastases become clinically significant secondary tumors, angiogenesis is essential for blood vessel development. A close relationship exists between Gal-3 and

the morphogenesis of endothelial cells and angiogenesis. 50-52 It was shown that Gal-3 behaves as a potent angiogenic factor through endothelial chemoattraction and cel1 motility induction, Matrigel invasion, capillary tube formation. 40,50 It was confirmed that MCP halted angiogenic activity of Gal-3. In a dosedependent way, MCP inhibited human endothelial cells' chemotaxis towards Gal-3, and it also inhibited endothelial cell capillary tube formation in vitro.<sup>50</sup> Further, it decreased spontaneous metastasis and angiogenesis when given to tumor-bearing mice.<sup>50</sup>

MCP can influence chemoresistance. Most anticancer agents work by inducing tumor cell apoptosis through the mitochondrial apoptosis pathway.<sup>53</sup> Gal-3 mitigates this pathway <sup>54</sup>. As a result, Gal-3 was demonstrated to directly modulate the cancer cell sensitivity to chemotherapeutic agents, for example, staurosporine,<sup>54</sup> cisplatin,<sup>54,55</sup> bortezomib,<sup>56</sup> etoposide,<sup>55</sup> doxorubicin,<sup>49</sup> and dexamethasone.<sup>56</sup> Thus, MCP as a Gal-3 blocker may

significantly affect the sensitivity of cancer cells to chemotherapy by limiting the anti-apoptotic effects of Gal-3 on the mitochondrial apoptosis of antipathway. The inhibition apoptotic effects of Gal-3 by MCP was shown to be adequate to augment the response of multiple myeloma cells to dexamethasone-induced apoptosis and their resistance to reverse to bortezomib.56 Upon MCP intervention hemangiosarcoma cells. their on sensitivity to doxorubicin-induced apoptosis was significantly elevated.<sup>49</sup>

It was demonstrated that MCP not only enhances anticancer drug-induced apoptosis but also induces cancer cell apoptosis by itself. MCP induced apoptosis via the caspase-8-to-caspase-3 pathway in multiple myeloma cells, notably without substantial alterations in mitochondrial membrane potential. <sup>56</sup>

The anticancer activity in different reports continues. A previous study reported that Gal-3 stimulated the activation of signal transducer and activator of transcription 3 (STAT3); its

constitutive activation in ovarian cancer cells is related to chemoresistance. Moreover, paclitaxel and **MCP** combination showed synergistic with decreased cytotoxicity viability and elevated caspase-3 activity in human SKOV-3 ovarian cancer cells.<sup>57</sup> Another study showed that MCP synergized with paclitaxel against SKOV-3 multicellular tumor spheroids by inhibiting STAT3 activation, decreasing its downstream target hypoxia-inducible factor- $1\alpha$  (HIF- $1\alpha$ ), lowering integrin mRNA levels, and consequently reducing protein kinase B activity.58

MCP enhanced the cytotoxicity of ionizing radiotherapy in a prostatic cell line (PCa) by decreasing anti-apoptotic Gal-3, elevating reactive oxygen species formation, and modulating DNA repair pathways. Moreover, MCP inhibited the PCa metastatic phenotype through Gal-3 blockade.<sup>59</sup> Furthermore, the MCP and doxorubicin combination resulted in progressive cytotoxicity in prostate cancer cell lines, leading to cell death mediated by cell cycle arrest in

LNCaP and apoptosis in DU-145.60 MCP-induced cytotoxicity of androgendependent and -independent prostate cancer cell lines may be partly due to mitogen-activated protein kinase signaling inhibition and caspase-3 activation.61 Furthermore. **MCP** synergistically diminished the invasive metastatic behavior of highly metastatic human prostate and breast cancer cells in vitro when combined with prostate or polybotanical health breast respectively.<sup>62</sup> supplements, Additionally, MCP-induced Gal-3 blockade inhibited tumor-associated macrophages (TAMs) induced breast cancer progression and metastasis in hypoxia; hypoxia elevates formation and secretion of Gal-3 from TAMs.63

It was evident that MCP decreased the growth of colon tumors that were implanted in mice.<sup>33</sup> In a previous *in vitro* study, MCP blocked extracellular Gal-3-induced human colon cancer cell migration at which Gal-3 binds to epidermal growth factor receptor, inducing colon cancer cell migration.<sup>64</sup>

Moreover, MCP inhibited liver metastasis in colon cancer in mice through Gal-3 inhibition.<sup>65</sup>

It has been shown that MCP-mediated Gal-3 inhibition decreased urinary bladder cancer proliferation and survival through apoptosis induction and cell cycle arrest *in vivo* and *in vitro*. <sup>66</sup>

In blood cultures, MCP significantly activated natural killer cells and Thelper cells. Additionally, natural killer cells exhibited functionality against K562 leukemic cells. It appears that the **MCP** immunostimulatory carbohydrates consist of low-Mw pectin polymer rich in unsaturated and saturated oligo-galacturonic acids, as well as a low degree of methyl esterification <sup>67</sup>. It is worth noting that MCP enhanced the cytotoxic activity of methotrexate both in choriocarcinoma acute (JEG3) and lymphoblastic leukemia (Nalm6) cell lines.<sup>68</sup>

The anticancer activity of MCP is affected by its size and domain

structures. Using autoclaving for MCP production, enriching de-esterified HG oligomers, and reducing RG-I and arabinogalactans-I (AG-I) in MCP lower than 3 KDa, or decreasing RG-I and increasing AG-I in MCP between 10-30 KDa led to anticancer activity by cancer cell inhibiting migration, proliferation, and aggregation. <sup>69</sup> MCP is ideal as an adjunctive immune and oncological therapy with its esterification degree, low Mw, and high RG-II domain percentage.<sup>70</sup>

Different clinical trials have demonstrated the potential anticancer effects of MCP. A phase II open-labeled pilot study assessed patients with prostate cancer, untreated at baseline, with low prostate-specific antigen (PSA) (<10 ng/ml) but gradually rising. MCP was administered daily for 12 months at a dose of 18 capsules (14.4 g). In 70% of patients, PSA doubling time was prolonged. These results suggest a slower progression of cancer and possible life extension.<sup>71</sup> An initial pilot trial examined 7 prostate cancer patients (PSA ranges 0.63-7.5) who had relapsed

or failed previous therapy. The daily MCP dose was 15 g. Four of seven patients had a positive response (>30% PSA doubling time prolongation), one had a stable state, one had a partial response, and one did not. Three years of survival were observed in all patients.<sup>72</sup> A phase II open-labeled assessing study non-metastatic biochemically relapsed prostate cancer in thirty-four patients who administered 4.8 g of MCP three times daily for 6 months. Six patients experienced grade 1 side effects (bloating and gas), but no patient experienced grade 3 or 4 toxicity. Stable or decreased PSA with negative scans was observed in 21 patients. Moreover, stable or improved PSA doubling time with no metastasis on scans was observed in 27 patients.<sup>73</sup> After this initial six months of MCP, a second long-term treatment for 12 months with patients exhibiting no disease progression was performed. After MCP therapy for 18 months, 90% (n=35) showed improved PSA doubling time, and all presented negative scans. No one exhibited grade 3 or 4 toxicity, and consequently, MCP exhibits sustained long-term safety and efficacy in biochemically relapsed prostate cancer.<sup>74</sup>

Patients suffering from solid tumors at an advanced stage were examined in an open-label clinical trial. Each treatment cycle consisted of eight weeks of daily administration of 15 g MCP. 20.7% (6 of 29 patients) experienced clinical

# 4. Protective effects of MCP on experimental and clinical studies

The protective effects of MCP were evidenced in several disease models, which are summarized in Table 1.

#### 4.1. Cardiovascular effects

A previous study reported that MCP ameliorated heart fibrosis elicited by isoproterenol in rats through inhibition of the Gal-3/TLR-4 (toll-like receptor-4)/NF-κB (nuclear factor kappa B) signaling pathway, thereby reducing the cardiac levels of proinflammatory cytokines such as tumor necrosis factor-

benefits and improved quality of life. After 2 cycles, 22.5% (11 of 49 patients) exhibited a stable disease, and 12.3% (6 of 49 patients) had a stable disease for over 24 weeks. One metastasized prostate cancer patient had a 50% reduction of serum PSA levels after sixteen weeks of therapy, along with enhanced quality of life, clinical benefit, and pain relief.<sup>75</sup>

alpha (TNF- $\alpha$ ), interleukin-18, and interleukin-1β (IL-1β), implicating in heart failure pathogenesis.<sup>76</sup> Moreover, MCP protected against isoproterenolcaused cardiac hypertrophy in rats through activation of p38 signaling and blocking Gal-3/TLR-4/JAK2 (Janus kinase 2)/STAT3 signaling pathway.<sup>77</sup> Recently, MCP showed protective effects against isoprenaline-caused myocardial infarction through Gal-3 inhibition, restoring echocardiographic parameters, and protection against remodeling.<sup>78</sup> Additionally, cardiac inhibition MCP-induced Gal-3 protected against isoproterenol-elicited left ventricular dysfunction and fibrosis.<sup>79</sup> MCP prevented cardiac

alterations related ischemic to reperfusion (IR) injury. It attenuated cardiac fibrosis and inflammation in the rats' left ventricles and protected against extracellular matrix remodeling through Gal-3 inhibition.<sup>80</sup> Also, MCP mitigated kidney fibrosis and heart dysfunction in an animal model of hyperaldosteronism through blockade.81 In hypertensive rats with hyperaldosteronism, MCP reversed vascular inflammation, hypertrophy, and fibrosis through Gal-3 inhibition.82 MCP-mediated Gal-3 inhibition protected against cardiac lipotoxicity in obese rats, identified by decreasing total triglycerides, lysophosphatidylcholine

#### 4.2.Renoprotective effects

The MCP's anti-apoptotic and antifibrotic effects were mediated through Gal-3 blocking in cisplatin-evoked kidney damage in mice.<sup>87</sup> It exhibited renoprotective effects in folic acidinduced acute renal damage in mice through Gal-3 inhibitory, antiapoptotic, anti-inflammatory, and antifibrotic impacts.<sup>88</sup> Further, in levels, and reactive oxygen species.83 Gal-3 inhibition MCP-caused facilitated the upregulation peroxiredoxin-4, thereby decreasing cardiac oxidative stress in doxorubicincaused cardiotoxicity in rats.84 As mentioned previously, MCP inhibited Gal-3/TLR-4/NF-κB signaling in a rat arteriogenic model of erectile dysfunction, reducing inflammation, fibrosis, and endothelial injury.85 Most recently, MCP protected against aortic dissection through inhibiting Gal-3/TLR-4 signaling and blocking pyroptotic macrophage-induced inflammation.86

spontaneously hypertensive rats, MCP had Gal-3 inhibitory, antifibrotic anti-inflammatory effects. and properties. It reduced the levels of inflammatory mediators. including Cd80, Cd68, Cd44, Cd45, osteopontin, chemoattractant protein-1, and addition to fibrotic markers, TGF-β, and collagen type I.89 Moreover, in two models of normotensive rats, MCP ameliorated mild renal injury induced by either aortic stenosis or obesity via

Gal-3 inhibition, which offered antiinflammatory and antifibrotic effects.<sup>90</sup> Recently, MCP ameliorated oxidative stress, fibrosis, inflammation, and apoptosis beyond its glycemic control against type-2 diabetes mellitus-elicited nephropathy in mice.<sup>74</sup>

#### 4.3. Hepatoprotective effects

By preventing fibrosis and apoptosis in rats exposed to carbon tetrachloride **MCP** (CCL4), demonstrated hepatoprotective effects via Gal-3 inhibitory effects.<sup>91</sup> It was recently demonstrated that MCP protects against methotrexate-caused liver and pulmonary damage in rats through antioxidant identified effects. by decreased malondialdehyde (MDA)

levels, increased superoxide dismutase (SOD) activity, nuclear factor erythroid 2 related-factor 2 (Nrf2), and reduced glutathione levels, anti-inflammatory effects, mediated by the blockade of Gal-3/TLR-4/NF-κB pathway, antifibrotic effects, mediated by Gal-3 inhibition, reduced collagen and TGF-β levels, and antiapoptotic effects.

#### 4.4. Neuroprotective effects

Through Gal-3 blockade. **MCP** protected against post-subarachnoid hemorrhage-induced disruption in the blood-brain barrier in mice, with mechanisms that may involve TLR-4 and extracellular signal-related kinase 1/2 (ERK 1/2), STAT3, and metalloproteinase-9 (MMP-9) in activation. 92 It was reported that *in vitro* and in vivo models of diabetes-induced cognitive impairment, MCP offered

antioxidant effects. indicated reduced levels of MDA and increased activity of SOD and glutathione peroxidase, and anti-inflammatory effects. demonstrated Gal-3 bv inhibition and decreased levels of proinflammatory cytokines such as TNF- $\alpha$ , IL-1 $\beta$ , and interleukin-6. 93 Previous experimental investigations uncovered the neuroprotective effects of MCP against ischemic stroke via

inhibiting Gal-3/TLR-4/NF- $\kappa$ B/NLRP-3 (NOD-like receptor 3)/cleaved caspase-1/IL-1 $\beta$  signaling pathway in microglia. 94 It mitigated cognitive

#### 4.5.Detoxifying effects

It was proved clinically that the urinary excretion of cadmium, arsenic, and lead is increased by MCP in healthy subjects. 96 In 5 case reports, MCP decreased levels of heavy metals with no side effects when administered alone or in combination with alginates. 97 In

#### 4.6.Miscellaneous effects

Of note, MCP had protective effects against articular cartilage defects in rabbits through Gal-3 inhibitory, antiinflammatory, and antidegenerative effects.<sup>99</sup> Further, it showed synergistic effects with hyaluronate against osteoarthritis in rabbits through modulating metabolic and inflammatory processes and consequently alleviating the progression of osteoarthritis. 100 Both MCP and Honokiol (HNK) showed deficits and neuroinflammation by blocking Gal-3 in addition to its antioxidant potential in scopolamineinduced Alzheimer's in rats.<sup>95</sup>

hospitalized children with lead toxicity, MCP safely decreased the serum level of lead and increased its level in urine. 10 MCP/alginate Furthermore. supplementation enhanced the excretion of uranium in feces with no side effects in a family exposed chronically to uranium in their environment and diet.<sup>98</sup>

antioxidant and anti-inflammatory effects when explored in vitro. MCP showed a higher antioxidant effect than HNK in a dose-dependent way. Of note, they demonstrated higher synergistic antioxidant and anti-inflammatory effects upon combination. This was evidenced inhibiting by lipid peroxidation, NF-κB, and cyclooxygenase-II.9 Additionally, the immunomodulatory effects of MCP were identified in the mouse spleen, elevated the levels. which of cytokines, proinflammatory which

could be advantageous in immunotherapy.<sup>11</sup>

Coadministration of MCP with probiotic supplement *Lactobacillus acidophilus* ATCC 4356 improved intestinal microbiota population and integrity. There was a significant elevation in fecal lactobacilli number in MCP alginate probiotic-challenged mice.<sup>101</sup> It was previously reported that

the adhesion of *Escherichia coli*, which produces toxins, and the cytotoxicity of Shiga toxin were decreased by MCP. <sup>102</sup> MCP exhibited antimicrobial activity against *Staphylococcus aureus* (MRSA) *in vitro*. In addition, it showed additives, with most of the MRSA strains, and synergetic, with two MRSA strains, effects when combined with cefotaxime. <sup>103</sup>

Table 1: Protective effects of modified citrus pectin on experimental and clinical studies

Effect	Disease model	Outcomes summary
Cardioprotective effect <sup>76</sup>	Isoproterenol-induced heart failure in rats	MCP inhibited the Gal-3/TLR-4/NF-κB signaling pathway. It decreased the expressions of IL-18, TNF-α, and IL-1β
Cardioprotective effect <sup>77</sup>	Isoproterenol-caused cardiac hypertrophy in rats	MCP activated p38 signaling and blocked Gal-3/TLR-4/JAK2 pathway
Cardioprotective effect <sup>78</sup>	Isoprenaline-caused myocardial infarction in type-2 diabetes mellitus rats	The echocardiographic parameters were restored with MCP. Further, it inhibited Gal-3 in parallel to its protective effect against cardiac remodeling

Cardioprotective effect <sup>79</sup>	Isoproterenol-elicited left ventricular dysfunction and fibrosis in mice	MCP inhibited Gal-3, which protected against isoproterenol-elicited left ventricular dysfunction and fibrosis
Cardioprotective effect <sup>80</sup>	Ischemia-reperfusion- induced myocardial injury in rats	MCP blocked Gal-3, concomitant with reduced cardiac inflammation and fibrosis. It decreased the ischemic area and extracellular matrix remodeling
Cardioprotective effect <sup>83</sup>	Cardiac lipotoxicity in high-fat diet-induced obesity in rats	MCP blocked Gal-3, which mediated a decrease in total triglycerides, phosphatidylcholine levels, and reactive oxygen species
Cardioprotective effect <sup>84</sup>	Doxorubicin-caused cardiotoxicity in rats	MCP upregulated peroxiredoxin-4 through Gal-3 inhibition and therefore decreased oxidative stress
Vascular protective effect <sup>82</sup>	Aldosterone-caused vascular fibrosis in rats	MCP reversed vascular inflammation, hypertrophy, and fibrosis, identified by reducing collagen type I content, through Gal-3 inhibition
Vascular protective effect <sup>85</sup>	High-fat diet-induced arteriogenic erectile dysfunction in rats	MCP inhibited Gal-3/TLR-4/NF-κB signaling. It decreased levels of TNF-α, IL-6, TGF-β, and collagen type I
Vascular protective effect <sup>86</sup>	β-aminopropionitrile fumarate/angiotensin-II induced aortic dissection in mice  RAW264.7 cells treated with H <sub>2</sub> O <sub>2</sub> ( <i>In vitro</i> )	In vivo, MCP decreased the mortality and incidence of aortic dissection. It reduced the infiltration of inflammatory cells into the aorta. Additionally, it blocked Gal-3/TLR4 signaling

		<i>In vitro</i> , MCP ameliorated pyroptosis and macrophage death induced by H <sub>2</sub> O <sub>2</sub>
Cardio-renoprotective effect <sup>81</sup>	Aldosterone-induced heart and kidney injuries in rats	MCP inhibited the aldosterone-induced increase in blood pressure, cardiac remodeling, and fibrosis. It lowered TGF-β and collagen type I. Also, it abrogated aldosterone-induced hyperfiltration, albuminuria, kidney and glomerular hypertrophy, tubular lesions, and renal fibrosis.
Renoprotective effect <sup>87</sup>	Cisplatin-evoked kidney damage in mice	MCP blocked renal Gal-3, which led to suppression of protein kinase α, apoptosis, cleaved caspase 3, collagen type I, and fibronectin.
Renoprotective effect <sup>88</sup>	Folic acid-induced acute renal damage in mice	MCP reduced renal proliferation in the acute phase of folic acid-induced acute renal damage. It decreased the levels of Gal-3, TGF- $\beta$ , collagen type I, fibronectin, $\alpha$ -SMA, IL-1 $\beta$ , and TNF- $\alpha$ in the injury recovery phase.
Renoprotective effect <sup>89</sup>	Spontaneously hypertensive rats	MCP inhibited Gal-3. It reduced the levels of inflammatory mediators, including Cd80, Cd68, Cd44, Cd45, osteopontin, and chemoattractant protein-1, in addition to fibrotic markers, TGF-β, and collagen type I.
Renoprotective effect <sup>90</sup>	Two models of renal damage in normotensive rats induced by either aortic stenosis or obesity	In the obesity model, MCP blocked Gal-3, decreased TGF-β and collagen type I as fibrotic markers, normalized α-SMA and E-cadherin as EMT markers, decreased osteopontin as an inflammatory marker,

		and ameliorated the kidney injury molecule 1. In the aortic stenosis model, MCP blocked Gal-3, decreased connective tissue growth factor, TGF- $\beta$ , and collagen type I, normalized $\alpha$ -SMA, E-cadherin, fibronectin, and $\beta$ -catenin, reduced osteopontin, and ameliorated the kidney injury molecule 1 and NGAL
Renoprotective effect <sup>74</sup>	Type-2 diabetes mellitus- elicited nephropathy in mice	MCP decreased MDA levels, elevated catalase activity, and GSH levels. It reduced iNOS, TGF-βRII, TNF-α, and caspase-3 levels.
Hepatoprotective effect <sup>91</sup>	Carbon tetrachloride- induced liver fibrosis	MCP inhibited Gal-3. It decreased fibrosis markers, α-SMA, TIMP-1, and collagen type I. Moreover, it reduced MDA levels and increased GSH content and SOD activity. It mediated the activation of hepatic stellate cells and apoptosis induction.
Hepato- and pulmonary protective effects <sup>68</sup>	Methotrexatecaused liver and pulmonary damage in rats	Both in liver and lung tissues, MCP decreased MDA levels, increased SOD activity, Nrf2, and GSH levels. It blocked Gal-3/TLR-4/NF-κB pathway and reduced collagen and TGF-β levels, and cleaved caspase-3
Neuroprotective effect <sup>92</sup>	Post-subarachnoid hemorrhage-induced disruption of the blood- brain barrier in mice	MCP inactivated TLR-4 and ERK 1/2, STAT3, and MMP9 through Gal-3 blockade.

Neuroprotective effect <sup>93</sup>	Diabetes-induced cognitive impairment <i>in vivo</i> in rats and <i>in vitro</i> in microglia cells	MCP blocked Gal-3, reduced MDA levels, increased activity of SOD and glutathione peroxidase, and decreased levels of proinflammatory cytokines such as TNF-α, IL-1β, and IL-6
Neuroprotective effect <sup>94</sup>	Cerebral ischemia reperfusion in mice as an in vivo model  Oxygen-glucose deprivation/ reoxygenation in microglia and neuronal cells as in vitro models	MCP ameliorated cerebral cortex cell injury and decreased infarct volume, cerebral water content, and scores of neurological deficits in mice. Moreover, it reduced apoptosis and increased cell viability in neuronal cells. It inhibited Gal-3/TLR-4/NF-κB/NLRP-3/cleaved caspase-1/IL-1β signaling pathway in microglia
Neuroprotective effect <sup>95</sup>	Scopolamine-induced Alzheimer's in rats	MCP decreased Gal-3, IL-6, and TNF-α, elevated brain-derived neurotrophic factor and SOD activity, and enhanced the memory performance.
Chondroprotective <sup>99</sup>	Articular cartilage defects in rabbits	MCP decreased levels of Gal-3, MMP13, IL-1β, Collagen 1A2 and inhibited cartilage degeneration
Chondroprotective <sup>100</sup>	Osteoarthritis in rabbits	MCP and hyaluronate combination alleviated the signs and symptoms of osteoarthritis, protected against the degeneration of articular cartilage, and reduced synovial inflammation.
Detoxifying effect <sup>96</sup>	Metal toxicity (clinical trial)	MCP increased the urinary excretion of cadmium, arsenic, and lead in healthy subjects

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Detoxifying effect <sup>97</sup>	Metal toxicity (clinical trial)	MCP decreased levels of heavy metals with no side effects when administered alone or in combination with alginates
Detoxifying effect <sup>10</sup>	Lead toxicity in hospitalized children	MCP safely decreased the serum level of lead and increased its level in urine
Detoxifying effect <sup>98</sup>	Low-level chronic exposure to uranium	MCP/alginate supplementation enhanced the excretion of uranium in feces with no side effects
Immune function <sup>9</sup>	Inflammation (in vitro)	The MCP and Honokiol combination demonstrated higher synergistic antioxidant and anti-inflammatory effects, inhibiting lipid peroxidation, NF-κB, and cyclooxygenase-II.
Immune function <sup>101</sup>	Probiotic	There was a significant elevation in fecal lactobacilli number in MCP alginate probiotic-challenged mice
Antimicrobial effect <sup>103</sup>	Staphylococcus aureus (In vitro)	MCP had antimicrobial activity alone or upon combination with cefotaxime, additive and synergistic effects, against <i>Staphylococcus aureus</i>

**Abbreviations**: α-SMA: Alpha smooth muscle actin, EMT: Epithelial-mesenchymal transition, ERK: Extracellular signal-related kinase, Gal-3: Galectin-3, GSH: reduced glutathione, IL: Interleukin, iNOS: inducible nitric oxide synthase, JAK2: Janus kinase 2, MCP: Modified citrus pectin, MDA: Malondialdehyde, MMP: Metalloproteinase, NF-κB: Nuclear factor kappa B, NGAL: Neutrophil gelatinase-associated lipocalin, NLRP-3:NOD-like receptor 3, Nrf2: Nuclear factor erythroid 2 related-factor 2, SOD: Superoxide dismutase, STAT3: Signal transducer and activator of transcription 3, TGF- $\beta$ RII: Tumor growth factor  $\beta$  receptor II, TGF- $\beta$ : tumor growth factor beta, TIMP-1:

Tissue inhibitor metalloproteinase 1, TLR-4: toll-like receptor-4, TNF-α: Tumor necrosis factor alpha.

#### 5. Conclusion

Several benefits of MCP have been highlighted in both preclinical and clinical studies. It showed antioxidant, antifibrotic, anti-inflammatory, and anti-apoptotic effects mediating its protective effects in different disease models. Moreover, its anticancer activity is documented. It inhibits tumor cell growth and metastasis. As a natural

Gal-3 inhibitor, MCP can be combined with other chemotherapeutic agents as a chemosensitizer or even to protect from chemotherapy-associated adverse effects, but these applications warrant clinical studies to evaluate its effectiveness, appropriate doses, and safety profile before clinical applications.

#### 6. Conflict of Interest

The authors have declared that no competing interests exist.

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### 8. Authorship contribution statement

Randa Ismail drafted the manuscript, while Gehan H. Heeba contributed to its conceptualization and reviewing. Heba A. Habib, Aliaa F. Anter critically reviewed the manuscript, and all approved its final version

#### 9. References

1. Liu X, Wang B, Tang S, et al. Modification, biological activity, applications, and future trends of citrus fiber as a functional component: A comprehensive

review. *International journal of biological macromolecules*. Jun 2024;269(Pt 1):131798. doi:10.1016/j.ijbiomac.2024.131798

- 2. Yue Y, Wang B, Xi W, et al. Modification methods, biological activities and applications of pectin: A review. *International journal of biological macromolecules*. Dec 31 2023;253(Pt 8):127523. doi:10.1016/j.ijbiomac.2023.127523
- 3. Baldwin JL, Shah AC. Pectin-induced occupational asthma. *Chest*. Dec 1993;104(6):1936-7. doi:10.1378/chest.104.6.1936b
- 4. Pienta KJ, Naik H, Akhtar A, et al. Inhibition of spontaneous metastasis in a rat prostate cancer model by oral administration of modified citrus pectin. *Journal of the National Cancer Institute*. Mar 1 1995;87(5):348-53. doi:10.1093/jnci/87.5.348
- 5. Morris VJ, Belshaw NJ, Waldron KW, Maxwell EG. The bioactivity of modified pectin fragments. *Bioactive Carbohydrates and Dietary Fibre*. 2013/01/01/ 2013;1(1):21-37. doi:https://doi.org/10.1016/j.bcdf.2013.02.001
- 6. Niture SK, Refai LJAJoP, Toxicology. Plant pectin: a potential source for cancer suppression. *American Journal of Pharmacology and Toxicology*. 2013;8(1):9.
- 7. Cao J, Yang J, Yue K, Wang Z. Preparation of modified citrus pectin (MCP) using an advanced oxidation process with hydroxyl radicals generated by UV-H2O2. *Food Hydrocolloids*. 2020/05/01/ 2020;102:105587. doi:https://doi.org/10.1016/j.foodhyd.2019.105587
- 8. Brouns F, Theuwissen E, Adam A, et al. Cholesterol-lowering properties of different pectin types in mildly hyper-cholesterolemic men and women. *European journal of clinical nutrition*. May 2012;66(5):591-9. doi:10.1038/ejcn.2011.208
- 9. Ramachandran C, Wilk B, Melnick SJ, Eliaz I. Synergistic Antioxidant and Anti-Inflammatory Effects between Modified Citrus Pectin and Honokiol. *Evidence-based complementary and alternative medicine : eCAM*. 2017;2017:8379843. doi:10.1155/2017/8379843
- 10. Zhao ZY, Liang L, Fan X, et al. The role of modified citrus pectin as an effective chelator of lead in children hospitalized with toxic lead levels. *Alternative therapies in health and medicine*. Jul-Aug 2008;14(4):34-8.

- 11. Merheb R, Abdel-Massih RM, Karam MC. Immunomodulatory effect of natural and modified Citrus pectin on cytokine levels in the spleen of BALB/c mice. *International journal of biological macromolecules*. 2019/01/01/2019;121:1-5. doi:https://doi.org/10.1016/j.ijbiomac.2018.09.189
- 12. Glinsky VV, Raz A. Modified citrus pectin anti-metastatic properties: one bullet, multiple targets. *Carbohydrate research*. Sep 28 2009;344(14):1788-91. doi:10.1016/j.carres.2008.08.038
- 13. Leclere L, Cutsem PV, Michiels C. Anti-cancer activities of pH- or heat-modified pectin. *Frontiers in pharmacology*. Oct 8 2013;4:128. doi:10.3389/fphar.2013.00128
- 14. Yang RY, Rabinovich GA, Liu FT. Galectins: structure, function and therapeutic potential. *Expert reviews in molecular medicine*. Jun 13 2008;10:e17. doi:10.1017/s1462399408000719
- 15. de Boer RA, van Veldhuisen DJ, Gansevoort RT, et al. The fibrosis marker galectin-3 and outcome in the general population. *Journal of internal medicine*. Jul 2012;272(1):55-64. doi:10.1111/j.1365-2796.2011.02476.x
- 16. Chiu YP, Sun YC, Qiu DC, et al. Liquid-liquid phase separation and extracellular multivalent interactions in the tale of galectin-3. *Nature communications*. Mar 6 2020;11(1):1229. doi:10.1038/s41467-020-15007-3
- 17. Zhao Z, Xu X, Cheng H, et al. Galectin-3 N-terminal tail prolines modulate cell activity and glycan-mediated oligomerization/phase separation. *Proceedings of the National Academy of Sciences of the United States of America*. May 11 2021;118(19)doi:10.1073/pnas.2021074118
- 18. Sundqvist M, Welin A, Elmwall J, et al. Galectin-3 type-C self-association on neutrophil surfaces; The carbohydrate recognition domain regulates cell function. *Journal of leukocyte biology*. Feb 2018;103(2):341-353. doi:10.1002/jlb.3a0317-110r
- 19. Liu FT, Stowell SR. The role of galectins in immunity and infection. *Nature reviews Immunology*. Aug 2023;23(8):479-494. doi:10.1038/s41577-022-00829-7
- 20. Mariño KV, Cagnoni AJ, Croci DO, Rabinovich GA. Targeting galectin-driven regulatory circuits in cancer and fibrosis. *Nature reviews Drug discovery*. Apr 2023;22(4):295-316. doi:10.1038/s41573-023-00636-2

- 21. Nangia-Makker P, Hogan V, Balan V, Raz A. Chimeric galectin-3 and collagens: Biomarkers and potential therapeutic targets in fibroproliferative diseases. *The Journal of biological chemistry*. Dec 2022;298(12):102622. doi:10.1016/j.jbc.2022.102622
- 22. Ahmed R, Anam K, Ahmed H. Development of Galectin-3 Targeting Drugs for Therapeutic Applications in Various Diseases. *International journal of molecular sciences*. May 1 2023;24(9)doi:10.3390/ijms24098116
- 23. Sciacchitano S, Lavra L, Morgante A, et al. Galectin-3: One Molecule for an Alphabet of Diseases, from A to Z. *International journal of molecular sciences*. Jan 26 2018;19(2)doi:10.3390/ijms19020379
- 24. Li LC, Li J, Gao J. Functions of galectin-3 and its role in fibrotic diseases. *The Journal of pharmacology and experimental therapeutics*. Nov 2014;351(2):336-43. doi:10.1124/jpet.114.218370
- 25. Hara A, Niwa M, Noguchi K, et al. Galectin-3 as a Next-Generation Biomarker for Detecting Early Stage of Various Diseases. *Biomolecules*. Mar 3 2020;10(3)doi:10.3390/biom10030389
- 26. Bouffette S, Botez I, De Ceuninck F. Targeting galectin-3 in inflammatory and fibrotic diseases. *Trends in pharmacological sciences*. Aug 2023;44(8):519-531. doi:10.1016/j.tips.2023.06.001
- 27. Nangia-Makker P, Hogan V, Raz A. Galectin-3 and cancer stemness. *Glycobiology*. Apr 1 2018;28(4):172-181. doi:10.1093/glycob/cwy001
- 28. Baldus SE, Zirbes TK, Weingarten M, et al. Increased galectin-3 expression in gastric cancer: correlations with histopathological subtypes, galactosylated antigens and tumor cell proliferation. *Tumour biology: the journal of the International Society for Oncodevelopmental Biology and Medicine*. Sep-Oct 2000;21(5):258-66. doi:10.1159/000030131
- 29. Radosavljevic G, Jovanovic I, Majstorovic I, et al. Deletion of galectin-3 in the host attenuates metastasis of murine melanoma by modulating tumor adhesion and NK cell activity. *Clinical & experimental metastasis*. Jun 2011;28(5):451-62. doi:10.1007/s10585-011-9383-y
- 30. Prieto VG, Mourad-Zeidan AA, Melnikova V, et al. Galectin-3 expression is associated with tumor progression and pattern of sun exposure in melanoma. Clinical cancer research: an official journal of the American Association for

Cancer Research. Nov 15 2006;12(22):6709-15. doi:10.1158/1078-0432.Ccr-06-0758

- 31. Clementy N, Piver E, Bisson A, et al. Galectin-3 in Atrial Fibrillation: Mechanisms and Therapeutic Implications. *International journal of molecular sciences*. Mar 25 2018;19(4)doi:10.3390/ijms19040976
- 32. Kim HR, Lin HM, Biliran H, Raz A. Cell cycle arrest and inhibition of anoikis by galectin-3 in human breast epithelial cells. *Cancer research*. Aug 15 1999;59(16):4148-54.
- 33. Hayashi A, Gillen AC, Lott JR. Effects of daily oral administration of quercetin chalcone and modified citrus pectin on implanted colon-25 tumor growth in Balb-c mice. *Alternative medicine review: a journal of clinical therapeutic.* Dec 2000;5(6):546-52.
- 34. Hsieh TC, Wu JM. Changes in cell growth, cyclin/kinase, endogenous phosphoproteins and nm23 gene expression in human prostatic JCA-1 cells treated with modified citrus pectin. *Biochemistry and molecular biology international*. Nov 1995;37(5):833-41.
- 35. Glinsky VV, Glinsky GV, Rittenhouse-Olson K, et al. The role of Thomsen-Friedenreich antigen in adhesion of human breast and prostate cancer cells to the endothelium. *Cancer research*. Jun 15 2001;61(12):4851-7.
- 36. Khaldoyanidi SK, Glinsky VV, Sikora L, et al. MDA-MB-435 human breast carcinoma cell homo- and heterotypic adhesion under flow conditions is mediated in part by Thomsen-Friedenreich antigen-galectin-3 interactions. *The Journal of biological chemistry*. Feb 7 2003;278(6):4127-34. doi:10.1074/jbc.M209590200
- 37. Glinsky VV, Glinsky GV, Glinskii OV, et al. Intravascular metastatic cancer cell homotypic aggregation at the sites of primary attachment to the endothelium. *Cancer research*. Jul 1 2003;63(13):3805-11.
- 38. Lehr JE, Pienta KJ. Preferential adhesion of prostate cancer cells to a human bone marrow endothelial cell line. *Journal of the National Cancer Institute*. Jan 21 1998;90(2):118-23. doi:10.1093/jnci/90.2.118
- 39. Platt D, Raz A. Modulation of the lung colonization of B16-F1 melanoma cells by citrus pectin. *Journal of the National Cancer Institute*. Mar 18 1992;84(6):438-42. doi:10.1093/jnci/84.6.438

- 40. Nangia-Makker P, Hogan V, Honjo Y, et al. Inhibition of human cancer cell growth and metastasis in nude mice by oral intake of modified citrus pectin. *Journal of the National Cancer Institute*. Dec 18 2002;94(24):1854-62. doi:10.1093/jnci/94.24.1854
- 41. Glinskii OV, Huxley VH, Glinsky GV, et al. Mechanical entrapment is insufficient and intercellular adhesion is essential for metastatic cell arrest in distant organs. *Neoplasia (New York, NY)*. May 2005;7(5):522-7. doi:10.1593/neo.04646
- 42. Al-Mehdi AB, Tozawa K, Fisher AB, et al. Intravascular origin of metastasis from the proliferation of endothelium-attached tumor cells: a new model for metastasis. *Nat Med*. Jan 2000;6(1):100-2. doi:10.1038/71429
- 43. Inohara H, Raz A. Effects of natural complex carbohydrate (citrus pectin) on murine melanoma cell properties related to galectin-3 functions. *Glycoconj J.* Dec 1994;11(6):527-32. doi:10.1007/bf00731303
- 44. Sathisha UV, Jayaram S, Harish Nayaka MA, Dharmesh SM. Inhibition of galectin-3 mediated cellular interactions by pectic polysaccharides from dietary sources. *Glycoconj J.* Nov 2007;24(8):497-507. doi:10.1007/s10719-007-9042-3
- 45. Chambers AF, Groom AC, MacDonald IC. Dissemination and growth of cancer cells in metastatic sites. *Nature reviews Cancer*. Aug 2002;2(8):563-72. doi:10.1038/nrc865
- 46. Nakahara S, Oka N, Raz A. On the role of galectin-3 in cancer apoptosis. *Apoptosis: an international journal on programmed cell death*. Mar 2005;10(2):267-75. doi:10.1007/s10495-005-0801-y
- 47. Yang RY, Liu FT. Galectins in cell growth and apoptosis. *Cellular and molecular life sciences: CMLS.* Feb 2003;60(2):267-76. doi:10.1007/s000180300022
- 48. Nangia-Makker P, Nakahara S, Hogan V, Raz A. Galectin-3 in apoptosis, a novel therapeutic target. *Journal of bioenergetics and biomembranes*. Feb 2007;39(1):79-84. doi:10.1007/s10863-006-9063-9
- 49. Johnson KD, Glinskii OV, Mossine VV, et al. Galectin-3 as a potential therapeutic target in tumors arising from malignant endothelia. *Neoplasia (New York, NY)*. Aug 2007;9(8):662-70. doi:10.1593/neo.07433

- 50. Nangia-Makker P, Honjo Y, Sarvis R, et al. Galectin-3 induces endothelial cell morphogenesis and angiogenesis. *The American journal of pathology*. Mar 2000;156(3):899-909. doi:10.1016/s0002-9440(10)64959-0
- 51. Fukushi J, Makagiansar IT, Stallcup WB. NG2 proteoglycan promotes endothelial cell motility and angiogenesis via engagement of galectin-3 and alpha3beta1 integrin. *Molecular biology of the cell*. Aug 2004;15(8):3580-90. doi:10.1091/mbc.e04-03-0236
- 52. Thijssen VL, Hulsmans S, Griffioen AW. The galectin profile of the endothelium: altered expression and localization in activated and tumor endothelial cells. *The American journal of pathology*. Feb 2008;172(2):545-53. doi:10.2353/ajpath.2008.070938
- 53. Pommier Y, Sordet O, Antony S, Hayward RL, Kohn KW. Apoptosis defects and chemotherapy resistance: molecular interaction maps and networks. *Oncogene*. Apr 12 2004;23(16):2934-49. doi:10.1038/sj.onc.1207515
- 54. Yu F, Finley RL, Jr., Raz A, Kim HR. Galectin-3 translocates to the perinuclear membranes and inhibits cytochrome c release from the mitochondria. A role for synexin in galectin-3 translocation. *The Journal of biological chemistry*. May 3 2002;277(18):15819-27. doi:10.1074/jbc.M200154200
- 55. Fukumori T, Oka N, Takenaka Y, et al. Galectin-3 regulates mitochondrial stability and antiapoptotic function in response to anticancer drug in prostate cancer. *Cancer research*. Mar 15 2006;66(6):3114-9. doi:10.1158/0008-5472.Can-05-3750
- 56. Chauhan D, Li G, Podar K, et al. A novel carbohydrate-based therapeutic GCS-100 overcomes bortezomib resistance and enhances dexamethasone-induced apoptosis in multiple myeloma cells. *Cancer research*. Sep 15 2005;65(18):8350-8. doi:10.1158/0008-5472.Can-05-0163
- 57. Hossein G, Keshavarz M, Ahmadi S, Naderi N. Synergistic effects of PectaSol-C modified citrus pectin an inhibitor of Galectin-3 and paclitaxel on apoptosis of human SKOV-3 ovarian cancer cells. *Asian Pacific journal of cancer prevention:* APJCP. 2013;14(12):7561-8. doi:10.7314/apjcp.2013.14.12.7561
- 58. Hossein G, Halvaei S, Heidarian Y, et al. Pectasol-C Modified Citrus Pectin targets Galectin-3-induced STAT3 activation and synergize paclitaxel cytotoxic

effect on ovarian cancer spheroids. *Cancer medicine*. Aug 2019;8(9):4315-4329. doi:10.1002/cam4.2334

- 59. Conti S, Vexler A, Hagoel L, et al. Modified Citrus Pectin as a Potential Sensitizer for Radiotherapy in Prostate Cancer. *Integrative cancer therapies*. Dec 2018;17(4):1225-1234. doi:10.1177/1534735418790382
- 60. Tehranian N, Sepehri H, Mehdipour P, et al. Combination effect of PectaSol and Doxorubicin on viability, cell cycle arrest and apoptosis in DU-145 and LNCaP prostate cancer cell lines. *Cell biology international*. Jul 2012;36(7):601-10. doi:10.1042/cbi20110309
- 61. Yan J, Katz A. PectaSol-C modified citrus pectin induces apoptosis and inhibition of proliferation in human and mouse androgen-dependent and-independent prostate cancer cells. *Integrative cancer therapies*. Jun 2010;9(2):197-203. doi:10.1177/1534735410369672
- 62. Jiang J, Eliaz I, Sliva D. Synergistic and additive effects of modified citrus pectin with two polybotanical compounds, in the suppression of invasive behavior of human breast and prostate cancer cells. *Integrative cancer therapies*. Mar 2013;12(2):145-52. doi:10.1177/1534735412442369
- 63. Wang L, Li YS, Yu LG, et al. Galectin-3 expression and secretion by tumor-associated macrophages in hypoxia promotes breast cancer progression. *Biochemical pharmacology*. Aug 2020;178:114113. doi:10.1016/j.bcp.2020.114113
- 64. Wu KL, Kuo CM, Huang EY, et al. Extracellular galectin-3 facilitates colon cancer cell migration and is related to the epidermal growth factor receptor. *American journal of translational research*. 2018;10(8):2402-2412.
- 65. Liu HY, Huang ZL, Yang GH, Lu WQ, Yu NR. Inhibitory effect of modified citrus pectin on liver metastases in a mouse colon cancer model. *World journal of gastroenterology*. Dec 28 2008;14(48):7386-91. doi:10.3748/wjg.14.7386
- 66. Fang T, Liu DD, Ning HM, et al. Modified citrus pectin inhibited bladder tumor growth through downregulation of galectin-3. *Acta pharmacologica Sinica*. Dec 2018;39(12):1885-1893. doi:10.1038/s41401-018-0004-z
- 67. Ramachandran C, Wilk BJ, Hotchkiss A, et al. Activation of human T-helper/inducer cell, T-cytotoxic cell, B-cell, and natural killer (NK)-cells and induction of natural killer cell activity against K562 chronic myeloid leukemia

cells with modified citrus pectin. *BMC complementary and alternative medicine*. Aug 4 2011;11:59. doi:10.1186/1472-6882-11-59

- 68. Ismail R, Habib HA, Anter AF, Amin A, Heeba GH. Modified citrus pectin ameliorates methotrexate-induced hepatic and pulmonary toxicity: role of Nrf2, galectin-3/TLR-4/NF-κB/TNF-α and TGF-β signaling pathways. *Frontiers in pharmacology*. 2025;16:1528978. doi:10.3389/fphar.2025.1528978
- 69. do Prado SBR, Shiga TM, Harazono Y, et al. Migration and proliferation of cancer cells in culture are differentially affected by molecular size of modified citrus pectin. *Carbohydrate polymers*. May 1 2019;211:141-151. doi:10.1016/j.carbpol.2019.02.010
- 70. Eliaz I, Raz A. Pleiotropic Effects of Modified Citrus Pectin. *Nutrients*. Nov 1 2019;11(11)doi:10.3390/nu11112619
- 71. Guess BW, Scholz MC, Strum SB, et al. Modified citrus pectin (MCP) increases the prostate-specific antigen doubling time in men with prostate cancer: a phase II pilot study. *Prostate cancer and prostatic diseases*. 2003;6(4):301-4. doi:10.1038/sj.pcan.4500679
- 72. Strum S, Scholz M, McDermed J, McCulloch M, Eliaz I. Modified citrus pectin slows PSA doubling time: A pilot clinical trial. 1999:
- 73. Keizman D, Frenkel MA, Peer A, et al. Effect of pectasol-c modified citrus pectin (P-MCP) treatment (tx) on PSA dynamics in non-metastatic biochemically relapsed prostate cancer (BRPC) patients (pts): Primary outcome analysis of a prospective phase II study. American Society of Clinical Oncology; 2019.
- 74. Mahmoud HM, Abdel-Razik AH, Elrehany MA, Othman EM, Bekhit AA. Modified Citrus Pectin (MCP) Confers a Renoprotective Effect on Early-Stage Nephropathy in Type-2 Diabetic Mice. *Chemistry & biodiversity*. Jul 2024;21(7):e202400104. doi:10.1002/cbdv.202400104
- 75. Azémar M, Hildenbrand B, Haering B, Heim ME, Unger CJCmO. Clinical benefit in patients with advanced solid tumors treated with modified citrus pectin: a prospective pilot study. *Clinical medicine Oncology*. 2007;1:CMO. S285. doi:10.4137/cmo.S285
- 76. Xu GR, Zhang C, Yang HX, et al. Modified citrus pectin ameliorates myocardial fibrosis and inflammation via suppressing galectin-3 and TLR4/MyD88/NF-κB signaling pathway. *Biomedicine & pharmacotherapy* =

Biomedecine & pharmacotherapie. Jun 2020;126:110071. doi:10.1016/j.biopha.2020.110071

- 77. Li Y, Zhou WW, Sun JH, et al. Modified citrus pectin prevents isoproterenol-induced cardiac hypertrophy associated with p38 signalling and TLR4/JAK/STAT3 pathway. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie*. Nov 2021;143:112178. doi:10.1016/j.biopha.2021.112178
- 78. Mahmoud MM, Hassan MM, Elsayed HE, et al. Protective effect of Galectin-3 inhibitor against cardiac remodelling in an isoprenaline-induced myocardial infarction in type 2 diabetes. *Archives of physiology and biochemistry*. Feb 2025;131(1):94-107. doi:10.1080/13813455.2024.2387710
- 79. Vergaro G, Prud'homme M, Fazal L, et al. Inhibition of Galectin-3 Pathway Prevents Isoproterenol-Induced Left Ventricular Dysfunction and Fibrosis in Mice. *Hypertension (Dallas, Tex: 1979)*. Mar 2016;67(3):606-12. doi:10.1161/hypertensionaha.115.06161
- 80. Ibarrola J, Matilla L, Martínez-Martínez E, et al. Myocardial Injury After Ischemia/Reperfusion Is Attenuated By Pharmacological Galectin-3 Inhibition. *Scientific reports*. Jul 3 2019;9(1):9607. doi:10.1038/s41598-019-46119-6
- 81. Calvier L, Martinez-Martinez E, Miana M, et al. The impact of galectin-3 inhibition on aldosterone-induced cardiac and renal injuries. *JACC Heart failure*. Jan 2015;3(1):59-67. doi:10.1016/j.jchf.2014.08.002
- 82. Calvier L, Miana M, Reboul P, et al. Galectin-3 mediates aldosterone-induced vascular fibrosis. *Arteriosclerosis*, thrombosis, and vascular biology. Jan 2013;33(1):67-75. doi:10.1161/atvbaha.112.300569
- 83. Marín-Royo G, Gallardo I, Martínez-Martínez E, et al. Inhibition of galectin-3 ameliorates the consequences of cardiac lipotoxicity in a rat model of dietinduced obesity. *Disease models & mechanisms*. Feb 5 2018;11(2)doi:10.1242/dmm.032086
- 84. Tian Y, Lv W, Lu C, et al. Galectin-3 inhibition attenuates doxorubicin-induced cardiac dysfunction by upregulating the expression of peroxiredoxin-4. *Canadian journal of physiology and pharmacology*. Oct 2020;98(10):700-707. doi:10.1139/cjpp-2019-0700

- 85. Wang G, Li R, Feng C, et al. Galectin-3 is involved in inflammation and fibrosis in arteriogenic erectile dysfunction via the TLR4/MyD88/NF-κB pathway. *Cell death discovery*. Feb 20 2024;10(1):92. doi:10.1038/s41420-024-01859-x
- 86. Shih CC, Lin WL, Chuu CP, et al. Modified Citrus Pectin protects Aortic Dissection Development Involving Macrophage Pyroptosis. *Archives of biochemistry and biophysics*. Apr 17 2025:110428. doi:10.1016/j.abb.2025.110428
- 87. Li HY, Yang S, Li JC, Feng JX. Galectin 3 inhibition attenuates renal injury progression in cisplatin-induced nephrotoxicity. *Bioscience reports*. Dec 21 2018;38(6)doi:10.1042/bsr20181803
- 88. Kolatsi-Joannou M, Price KL, Winyard PJ, Long DA. Modified citrus pectin reduces galectin-3 expression and disease severity in experimental acute kidney injury. *PloS one*. Apr 8 2011;6(4):e18683. doi:10.1371/journal.pone.0018683
- 89. Martínez-Martínez E, Ibarrola J, Fernández-Celis A, et al. Galectin-3 pharmacological inhibition attenuates early renal damage in spontaneously hypertensive rats. *Journal of hypertension*. Feb 2018;36(2):368-376. doi:10.1097/hjh.0000000000001545
- 90. Martinez-Martinez E, Ibarrola J, Calvier L, et al. Galectin-3 Blockade Reduces Renal Fibrosis in Two Normotensive Experimental Models of Renal Damage. *PloS one*. 2016;11(11):e0166272. doi:10.1371/journal.pone.0166272
- 91. Abu-Elsaad NM, Elkashef WF. Modified citrus pectin stops progression of liver fibrosis by inhibiting galectin-3 and inducing apoptosis of stellate cells. *Canadian journal of physiology and pharmacology*. May 2016;94(5):554-62. doi:10.1139/cjpp-2015-0284
- 92. Nishikawa H, Liu L, Nakano F, et al. Modified Citrus Pectin Prevents Blood-Brain Barrier Disruption in Mouse Subarachnoid Hemorrhage by Inhibiting Galectin-3. *Stroke*. Nov 2018;49(11):2743-2751. doi:10.1161/strokeaha.118.021757
- 93. Yin Q, Chen J, Ma S, et al. Pharmacological Inhibition of Galectin-3 Ameliorates Diabetes-Associated Cognitive Impairment, Oxidative Stress and Neuroinflammation in vivo and in vitro. *Journal of inflammation research*. 2020;13:533-542. doi:10.2147/jir.S273858

- 94. Cui Y, Zhang NN, Wang D, Meng WH, Chen HS. Modified Citrus Pectin Alleviates Cerebral Ischemia/Reperfusion Injury by Inhibiting NLRP3 Inflammasome Activation via TLR4/NF-kB Signaling Pathway in Microglia. *Journal of inflammation research*. 2022;15:3369-3385. doi:10.2147/jir.S366927
- 95. Akgöl J, Kutlay Ö, Keskin Aktan A, Fırat F. Assessment of Modified Citrus Pectin's Effects on Dementia in the Scopolamine-Induced Alzheimer's Model in Adult Male Wistar Rats. *Current issues in molecular biology*. Dec 11 2024;46(12):13922-13936. doi:10.3390/cimb46120832
- 96. Eliaz I, Hotchkiss AT, Fishman ML, Rode D. The effect of modified citrus pectin on urinary excretion of toxic elements. *Phytotherapy research: PTR*. Oct 2006;20(10):859-64. doi:10.1002/ptr.1953
- 97. Eliaz I, Weil E, Wilk B. Integrative medicine and the role of modified citrus pectin/alginates in heavy metal chelation and detoxification--five case reports. *Forschende Komplementarmedizin (2006)*. Dec 2007;14(6):358-64. doi:10.1159/000109829
- 98. Eliaz I, Weil E, Schwarzbach J, Wilk B. Modified Citrus Pectin / Alginate Dietary Supplement Increased Fecal Excretion of Uranium: A Family. *Alternative therapies in health and medicine*. Jul 2019;25(4):20-24.
- 99. Zhang Y, Su D, Wang Y, et al. Locally delivered modified citrus pectin a galectin-3 inhibitor shows expected anti-inflammatory and unexpected regeneration-promoting effects on repair of articular cartilage defect. *Biomaterials*. Dec 2022;291:121870. doi:10.1016/j.biomaterials.2022.121870
- 100. Chen Y, Su D, Zheng J, et al. Intra-articular injection of modified citrus pectin and hyaluronate gel induces synergistic effects in treating osteoarthritis. *International journal of biological macromolecules*. Sep 2024;276(Pt 1):133840. doi:10.1016/j.ijbiomac.2024.133840
- 101. Odun-Ayo F, Mellem J, Reddy LJFS, Technology. The effect of modified citrus pectin-probiotic on faecal lactobacilli in Balb/c mice. *Food Science and Technology*. 2017;37(3):478-482.
- 102. Di R, Vakkalanka MS, Onumpai C, et al. Pectic oligosaccharide structure-function relationships: Prebiotics, inhibitors of Escherichia coli O157:H7 adhesion and reduction of Shiga toxin cytotoxicity in HT29 cells. *Food chemistry*. Jul 15 2017;227:245-254. doi:10.1016/j.foodchem.2017.01.100

103. Dahdouh E, El-Khatib S, Baydoun E, Abdel-Massih RM. Additive Effect of MCP in Combination with Cefotaxime Against Staphylococcus aureus. *Medicinal chemistry (Shariqah (United Arab Emirates))*. 2017;13(7):682-688. doi:10.2174/1573406413666170306112444